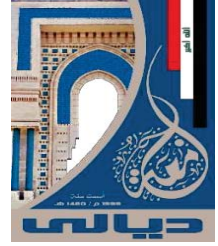


Republic of Iraq  
Ministry of Higher Education  
and Scientific Research  
University of Diyala  
College of Medicine



**Gene Detection of Some Virulence Factors In *Klebsiella  
Pneumoniae* Isolated From UTI**

A Thesis

Submitted to The Council College of Medicine - University of  
Diyala in Partial Fulfillment of the Requirements for the Master  
Degree in Medical Microbiology

**By**

**Sara Hadi Jassim**

BSc.of Science in Biology - University of Diyala

2016-2017

**Supervised by**

**Assistant Professor**

**Dr. Anfal Shakir Motib**

2020 A.D.

1442 A.H.

بِسْمِ اللَّهِ الرَّحْمَنِ الرَّحِيمِ

﴿ يُؤْتِي الْحِكْمَةَ مَنْ يَشَاءُ وَمَنْ يُؤْتَ الْحِكْمَةَ  
فَقَدْ أُوتِيَ خَيْرًا كَثِيرًا وَمَا يَذَّكَّرُ إِلَّا أُولُو  
الْأَلْبَابِ ﴾ ﴿٣٦٩﴾

صَدَقَ اللَّهُ الْعَظِيمُ<sup>1</sup>

# Dedication

To My Parents

For toughing me to trust in Allah, all the unconditional love, guidance, raised me to be the person I am today, supporting and encouraging me to believe in myself

My Husband

For his remarkable patience and unwavering love and support over the many years we have been together

My Brothers and Sister

For have been great sources of motivation, love and inspiration for me

*Sara*

## Acknowledgements

At First of all, my great thanks to Allah for enabling me to finish what I started and for helping me to present this work. The Prayers and the peace are upon our Prophet the master of the creatures Mohammed, and his genial modest family and his companions.

I would like to express my deep gratitude and sincere thanks to my supervisors **Dr. Anfal Shakir Motib** for her continuous support, valuable advices, constructive criticism and providing all required facilities to perform this work.

My thanks to the head and staff of College of Medicine/ University of Diyala.

I also indebted to staff of Baladroz hospital specially Ali Salah and Ali Hassim and Al-Batool hospital especially Ahmeed and Mustafa.

Also, Plentitude and Grate full thank to staff of Central Public Laboratory Health in Department of Bacteriology especially, Ms. Manal for the facilities that supplied to carry out this work.

I should not forget my private feeling of thankfulness to my parents, Hadi Jassim and Madhea, my sisters Dyana, Shahad and Noor and my brother (Hassan), and to my another family my husband's father (Nazar Hussein) and to my husband's mother (Zanab Hameed) and my husband's sister (Naba 'a) without their patience, helps and support, I could never have completed this thesis. Finally, the words cannot express my great love, thankfulness and heartfelt gratitude to my Husband (Haider Nazar Hussein) for his love, patience, support and remarkable aids during the period of the research.

*Sara*

## *Supervisor Certification*

We, Certify that this thesis entitled (**Gene Detection of Some Virulence Factors In *Klebsiella Pneumoniae* Isolated From UTI**) has been conducted under our supervision at College of Medicine, University of Diyala, as a partial requirements for the Master Degree in Medical Microbiology.

**Assist.Professor**

**Dr. Anfal Shakir Motib**

In view of available recommendation, I forward this thesis for debate by the examining committee.

**Signature**

**Assistant Professor Dr. Luma Taha Ahmed**

Head of Microbiology Department

College of Medicine – University of Diyala

## ***Examination Committee Certification***

We, the examining committee, certify that we have read this thesis entitled (**Gene Detection of Some Virulence Factors In *Klebsiella Pneumoniae* Isolated From UTI**) which prepared by (**Sara Hadi Jassim**) and have examined the student in its contents, and that in our opinion it is adequate for awarding the Degree of Master in Medical Microbiology.

**Signature:**

**Name: Dr. Karim Ibrahim Mubarak**

**Degree: Assist. Professor (Chairman)**

**Date: / /2020**

**Signature:**

**Signature:**

**Name: Dr. Jabbar Salman Hassan**

**Scientific Degree: Assist. Professor  
(Member)**

**Date: / /2020**

**Name: Dr. Hind Hussein Obaid**

**Scientific Degree: Assist. Professor  
(Member)**

**Date: / /2020**

**Signature:**

**Name: Dr. Anfal Shakir Motib**

**Scientific Degree: Assist. Professor  
(Member/Supervisor)**

**Date: / /2020**

**Approved by the Council of the College of Medicine – University of Diyala.**

**Signature:**

**Professor Dr. Ismail Ibrahim Latif  
Dean College of Medicine-University of Diyala**

**Date: / /2020**

## ***Abstract***

---

### **Abstract**

A total of 160 clinical samples were collected from two hospitals in Diyala from patients in both sexes, suffering from urinary tract infection of age range between ( $\leq$  10-50) years in the period between (September / 2019 –December / 2019). Out of 160 isolates, 24 identified as *Klebsiella pneumoniae* depending on cultural, biochemical characteristics, and molecular methods. The remaining bacterial isolates from urine samples were identified as *Escherichia coli* (45) , Pseudomonas(23), Proteus(27 and Citrobacter(16) isolates.

Regarding to the patients gender, it was found that females had a tendency to get infection more than the males when 20 (83.3%) of patients were females and 4 (16.7 %) of them are males. Moreover, the age group (30-39) were most subjected to the infection of *K. Pneumoniae*.

The antibiotic susceptibility test showed that all isolates were Multidrug Resistance (MDR) , were high resistant to Ciprofloxacin 24 (100%),Cefotaxime 24(100%) ,Piperacillin 17(70%), Nalidixic acid 12(50%), Tetracycline 10 (41%), Nitrofurantoin8(33%), Chloramphenicol 7(29%) Ampicillin-sulbactam7(29%), Tobromycin4(16%), and Imipenem 2(8%).

The organism were identified by using molecular methods, including the DNA extraction and checking with PCR using identified genes, including *16srRNA*,and some virulence factor gene such as *rmpA* and *iroN* gene .

The DNA sequencing was done to identify the strains of *Klebsiella pneumoniae* that cause UTI in Iraq especially in Diyala city, the results showed that there are two strains responsible to the UTI in this city, *Klebsiella pneumoniae* strains D16KP0042 and KPWIQ25. The results of DNA sequencing analysis of *16SrRNA* gene from k1 to k24 isolates revealed that

## ***Abstract***

---

*k.pneumoniae* were (100%).and iro N gene revealed only in 3 isolates(12.5%),while rmp A gene not observed in any isolates.

The prevalence of biofilm producer bacterial isolates was 22(91%) that 7(31%) produced strong, 6(27%) produced moderate and 9(40%) produced weakly biofilm.



## Table of Contents

Subject	Page No.
Dedication	
Acknowledgment	
Abstract	I
Table of Contents	III
List of Tables	VII
List of Figures	VIII
List of Abbreviations	X

Items	Subject	Page No.
Chapter One : Introduction		
1-1	Introduction	1
1-2	Aims of study	2
Chapter Two : Literatures review		
2.1	General description of <i>Klebsiella pneumoniae</i>	3
2.2	Epidemiology of <i>Klebsiella.Pneumoniae</i>	4
2.3	Virulence Factors of <i>Klebsiella</i>	6
2.4	Antibiotic resistance <i>Klebsiella. Pneumoniae</i> population	10
2.5	Urinary tract infection with <i>Klebsiella pneumoniae</i>	12
2.6	Biofilm Formation	14
2.6.1	The stages of biofilm formation	15
2.6.2	Factors play a role in biofilm formation	17
2.7	Correlation between Biofilm and Antibiotic Resistance	19
Chapter Three : Patients , Materials and Methods		

3.1	Patients	21
3.1.1	Collection of urine samples	21
3.1.2	Isolation of bacteria	21
3.2	Materials	22
3.2.1	Laboratory equipment	22
3.2.2	Tools	23
3.2.3	Culture media	24
3.2.4	Antibiotics supplied by a Bioanalyse company (Turkey)	24
3.2.5	Chemical materials	25
3.2.6	Kits, Primers	26
3.2.6.1	Kits	26
3.2.6.2	Primers	26
3.2.7	Preparation of Reagents and Solutions	27
3.2.7.1	Reagents	27
3.2.7.2	Solutions	27
3.3	Methods	28
3.3.1	Sterilization Methods	28
3.3.2	Preparation of culture media	29
3.3.3	Biochemical tests	29
3.3.4	Preservation and Maintenance of Bacterial Isolates	31
3.3.5	Antimicrobial Susceptibility Testing	31
3.3.6	Molecular methods for <i>Klebsiella Pneumoniae</i> detection	32
3.3.6.1	DNA Extraction	32
3.3.6.2	Quantification of DNA	33
3.3.7	PCR study	34
3.3.7.1	Detection of 16S r RNA gene of <i>klebsiella pneumoniae</i>	34
3.3.7.1.1	Primer preparation(16 S Primers)	34

3.3.7.1.2	Reaction Setup and Thermal Cycling Protocol	34
3.3.7.2	Detection of rmpA gene and ironN gene of <i>klebsiella pneumoniae</i>	36
3.3.8	Agarose Gel Electrophoresis	37
3.3.8.1	Solutions	37
3.3.8.2	Preparation of agarose gel	37
3.3.8.3	Casting of the horizontal agarose gel	38
3.3.8.4	DNA loading	38
3.3.9	Standard Sequencing	38
3.3.10	Effect of antibiotics on biofilm formation ability of isolates	38
3.3.11	Statistical analysis	40
Chapter four : Result		
4.1	Identification of <i>Klebsiella</i> isolates	41
4.2	Diagnostic methods of <i>Klebsiella pneumonia</i>	42
4.2.1	Bacteriology culture test	42
4.2.2	Microscopy examination	42
4.2.3	Biochemical test	42
4.3	Antibiotic susceptibility of <i>Klebsiella pneumonia</i>	44
4.4	Detection Molecular methods for <i>K.pneumoniae</i>	46

4.4.1	Detection of 16SrRNA gene of <i>K.pneumoniae</i>	46
4.4.2	Determination of <i>k.pneumoniae</i> Virulence Genes by PCR	46
4.4.3	DNA Sequencing	48
4.5	Distribution of <i>K. pneumoniae</i> according to gender and ages of patients	49
4.6	Comorbidities diseases associated with <i>Klebsiella pneumoniae</i> infected.	50
4.7	Biofilm formation assay	50
Chapter FIVE : DISCUSSION		
5.1	Isolation of <i>Klebsiella</i>	52
5.2	Culture characters	52
5.3	Biochemical test	53
5.4	Antimicrobial Susceptibility testing for each isolate	54
5.5	Molecular Detection for <i>K. pneumoniae</i>	56
5.6	The distribution of <i>Klebsiella</i> among individuals according to sex and age groups	58
5.7	Comorbidities diseases associated with <i>Klebsiella pneumoniae</i> infected.	59
5.8	Biofilm formation	60
Chapter SIX : Conclusions		
6.1.	Conclusions	62
6.2.	Recommendations	63
References		
	References	64
Appendix		
	Appendix (1)	

## List of Tables

<b>Table No.</b>	<b>Title</b>	<b>Page No.</b>
3-1	Common apparatus & equipment used in the present work	22
3-2	The general tools used in the present study.	23
3-3	Microbial culture media utilized for completion of study	24
3-4	Antibiotics used in the current study	24
3-5	Chemical compounds referred and utilized in the present work.	25
3-6	Following items were used in this study.	26
3-7	Primer used in the current study	26
3-8	The solution used in preparation of primers	34
3-9	PCR Component Calculation for 16srRNA primers	34
3-10	Monoplex PCR master mix	35
3-11	Program of PCR thermocycling conditions	35
3-12	Conditions used to amplify the <i>klebsiella pneumoniae</i> rmp A gene	36
3-13	Conditions used to amplify the <i>klebsiella pneumoniae</i> iro N gene.	37
3-14	Classification of bacterial adherent by tissue culture plate method	39
4-1	The results of some biochemical test of <i>Klebsiella pneumoniae</i> and others bacteria.	43
4-2	Sensitive, intermediate and resistance of <i>K.pneumoniae</i> isolates are compared by using X <sup>2</sup> test.	45

## List of Figures

<b>Figure No.</b>	<b>Title</b>	<b>Page No.</b>
2-1	Graphic illustrating the biofilm formation process	16
4-1	Frequency and percentage of microorganism isolated from urine samples of patients with UTI, p=0.003**	41
4-2	Agarose gel electrophoresis of DNA from urin sample directly showing PCR products for 1500 bp of 16s rRNA gene when compared to the molecular ladder (1500-100). The product was electrophoresis on 1% agarose at 5 volt/cm <sup>2</sup> for 1:30 hours.	46
4-3	Agarose gel electrophoresis of DNA from urin sample directly showing PCR products for 418 bp of rmpA gene when compared to the molecular ladder (1500-100). The product was electrophoresis on 1% agarose at 5 volt/cm <sup>2</sup> for 1:30 hours.	47
4-4	Agarose gel electrophoresis of DNA from urin sample directly showing PCR products for 556 bp of iro N gene when compared to the molecular ladder (1500-100). The product was electrophoresis on 1% agarose at 5 volt/cm <sup>2</sup> for 1:30 hours.	48
4-5	Distribution of patients according to gender.	49
4-6	Distribution of patients according to age periods.	49
4-7	Frequency and percentage of comorbidities diseases associated with	50
4-8	Biofilm formation assay of isolated microorganism values are the mean of triplicates $\pm$ SD where (p $\leq$ 0.005)	51

## List of Abbreviations

Abbreviate	Key
AMP	Antimicrobial Peptides
BaCl <sub>2</sub>	Barium chloride
C3b	Complement protein
CFU	Colony forming units
CPS	Polysaccharides of Capsule
CRBSIs	Catheter-related blood stream infections
CRKP	Carbapenem-resistant <i>Klebsiella pneumoniae</i>
D.W.	Distilled water
ESBL	Extended spectrum beta-lactamases
HAP	Hospital acquired pneumoniae
ICU	Intensive care unit
<i>K. pneumoniae</i>	<i>Klebsiella pneumoniae</i>
LPS	Lipopolysaccharide
MDR	Multidrug-resistance
PBS	Phosphate Buffer Saline
PCR	Polymerase Chain Reaction
<i>RmpA</i>	Regulator of phenotype gene A
rRNA	Ribosomal Ribonucleic Acid
SOD	Superoxide dismutase
UTI	Urinary tract infection
VP1-VP2	Vogas-Proskauer

# Chapter One

## Introduction



**Introduction.**

*Klebsiella pneumoniae* is a Gram-negative rod in the Enterobacteriaceae family. The bacterium is found indigenously in soil and waters, but also on mucosal surfaces in mammals, including humans (Podschun and Ullmann, 1998).

*Klebsiella pneumoniae* is responsible for many community –onset and nosocomial infections. The increasingly high level of antimicrobial drug resistance prevalence is an exaggerated problem, especially for healthcare providers. These bacterium can confer resistance to the majority of antibiotics by applying vast amounts of resistance mechanisms, leading to high mortality and morbidity rates. The dominant antibiotics used for treating this bacterium infections today are the  $\beta$ -lactam antibiotics, which inhibit transpeptidases participating in bacterial cell wall synthesis. Beta-lactam antibiotics can be deactivated by  $\beta$ -lactamase enzymes (Paterson and Bonomo, 2005).

Antibiotic resistance represents one of most significant healthcare problems. The loss of effective antibiotics would weaken the ability to fight infectious diseases and treat the complications for patients with renal dialysis, cancer patients with chemotherapy, and organ transplantation surgery (Organization, 2014).

The choice of an adequate antibiotic regimen for the treatment of infections due to Multidrug resistant (MDR) strains of *K. pneumoniae* is a challenge for physicians for the patients which develop serious infections which is from patients hospitalized considering that patients in many cases develop serious infections and complications , these infections affect not only critically ill patients hospitalized in intensive care unit (ICU) but also patients from other wards with multiple comorbidities(Bassetti et al., 2017) . The important and recognizable virulence factors and quorum sensing (QS) are involved in higher degree of infection in *Klebsiella pneumoniae*. QS is responsible for change in community of bacteria for their mobility alteration, different enzymes production which may help them to

invade and establish in tissues of host, toxic compound production, Slime and capsule components production and formation of aggregates in the form of flocks and biofilm (Papenfort and Bassler, 2016).

Biofilm formation with the help of QS can be recognized as intrinsic antibiotic resistance factor in this type of aggregates makes impermeability in a structure and stop antibiotic to interact with a bacteria and increased the antibiotic resistance (Vuotto et al., 2014). Therefore, it is essential to study the relationship between antibiotic resistance and biofilm formation in *K. pneumoniae*.

**1-2 Aim of study :**

1. To identify *k. pneumoniae* isolated from urinary tract infection patients using 16 S gene sequencing.
2. To find the antibiotic sensitivity of *K. pneumoniae* strains.
3. To investigated the rate of occurrence of biofilm in *k. pneumoniae* isolated from UTI.
4. To identify the correlation with different parameters such as gender, age, and chronic disease