

**Republic of Iraq  
Ministry of Higher Education  
and Scientific Research  
University of Diyala  
College of Medicine**



## **Association of human papillomavirus type 11, 16 and 18 among female patients with breast cancer in Baghdad.**

### **A Thesis**

Submitted to the Council of the College of Medicine - University of Diyala in Partial Fulfillment of the Requirements of the Degree of Master's of Sciences in Medical microbiology

**By**  
**Alaa Wa'el Izzat Al-Bekri**

**B.Sc. in Biology (2000) - College of Science - University of Baghdad**

**Supervised by**

**Professor Dr.  
Abdulrazak Shafiq Hasan  
Ph.D. in Medical Microbiology/virology**

**2021 A.D.**

**1442 A.H.**

بِسْمِ اللَّهِ الرَّحْمَنِ الرَّحِيمِ

الرَّحْمَنُ (١) عَلِمَ الْقُرْآنَ (٢) خَلَقَ الْإِنْسَانَ (٣) عَلِمَهُ  
البَيَانَ (٤)

صَدَقَ اللَّهُ الْعَظِيمُ

سورة الرحمن - الآيات (١ - ٤)

## **Supervisor Certification**

I, certify that this thesis entitled (**Association of human papillomavirus type 11,16 and 18 among female patients with breast cancer in Baghdad**) . has been conducted under my supervision at the College of Medicine, University of Diyala, as partial fulfilment of the requirements for the Master Degree of Science in Medical Microbiology.

**Professor Dr. Abdulrazak Shafiq Hasan**

Given the available recommendation, I forward this thesis for debate by the examining committee.

**Signature**

**Professor Dr. Luma Taha Ahmed**

Head of Microbiology Department

College of Medicine - University of Diyala

## **Committee Certification**

We, as the examining committee, certify that we have read this thesis and examined the student (Alaa Wael Izzat) in and its contents, found it adequate as a thesis for the Master Degree of Science in Medical Microbiology.

Professor

**Dr. Saad Hasan Mohammed Ali**

Chairman

Professor

Assistant Professor

**Dr. Nadhim Ghazal Noaman**

**Dr. Ahmed Majeed Al-Shammary**

Member

Member

**Professor**

**Dr. Abdulrazak Shafiq Hasan**

**(Supervisor)**

Approved by the Council of College of Medicine

The Dean

**Professor Dr. Ismail Ibrahim Latif**

Date:

## ***Dedication***

*To the soul of my brother, Ahmed Wa'el Al-Bekri, the Immaculate Martyr.*

*My dear mother, thank you for your love, support and everything you have gone through for me. I'm blessed because of you.*

*My dear father, Dr Wa'el Izzat Al-Bekri; my pride and happiness come from you.*

*Ahmed Ghanim Al-Hayali, my husband and soul mate, my lifelong friend and first supporter in the world.*

*To my treasure in both adversity and prosperity, Asal, Laya, and Ibrahim.*

*Without you all, my pen would not be able to write these words.*

## **Acknowledgement**

*I am grateful to Allah Almighty for the well-being and for inspiring me to be patient and determined to complete this study.*

*I would like to utter my sincere appreciation and thanks to my advisor Professor Dr Abdulrazak Shafiq Hasan, for suggesting the study proposal and for providing me with invaluable advice and knowledge throughout the study.*

*Special thanks to the pathologist Dr Areej Mustafa Kamal in the Cytological Examination department for her kind and immense support; and all the staff members of the laboratories in the Oncology Teaching Hospital at Baghdad Medical City and the staff of the Blood bank of the Nursing House Private Hospital and everyone who helped me.*

*I particularly thank Professor Dr Waleed Tawfiq Al-Ani, College of Medicine, University of Al-Mustansiriyah, Baghdad- Iraq, for his accurate statistical analysis of the data of the current study.*

*Thanks are due to Ms Hiba Al-Zuhairi at the Post Graduate Students' Department, at the College of Medicine – University of Diyala, for her unlimited support.*

*I would like to thank all my teachers and instructors who have taught me at all levels of my studies.*

*Finally, I am grateful to each patient woman who participated in this study and gave me her time and trust despite her pain and misery.*

## SUMMARY

Breast carcinoma is the commonest malignancy among women in developed and developing countries including Iraq. It ranked as the number one cancer documented in all Iraqi provinces. Breast cancer is one of the main causes of death in postmenopausal women, accounting for %23 of all cancer fatalities. According to Globocan registry data in 2020, there are 7515 persons, estimated 22.2% breast cancer registry from both genders from all ages with mortality of 3019 of Iraqi people. Previous studies have mentioned the carcinogenic role of HPV in transformed HPV virus-infected cells into a malignant phenotype, which is an important cause of cancer in humans.

This is a case-control study, which was conducted from August 2020 to July 2021 in an Oncology Teaching Hospital-Medical City and the Blood bank of the Private Nursing Home Hospital. This study enrolled 90 participants: 29 women apparently healthy as a control group and 61 women with clinically and laboratory diagnosed breast cancer as patients' group. Age ranges from 30 to 78 years old. A questionnaire form was made for this purpose. Blood samples were collected from each participant, Complete Blood Count was determined and a blood group was identified for each participant. Sera were separated and kept at -30° C till use. ELISA kits from SUNLONG company- China were used to detect the presence of HPV type 11,16,18 antigens and to determine overexpression of P53 tumour suppressor protein. Human privacy was respected by obtaining the verbal consent of the participants. Statistical analysis was done via SPSS version 27 and P -values less than 0.05 were considered significant.

All breast cancers were histologically diagnosed with almost advanced stages. The three types of HPV were identified in sera of examined Iraqi women with BC and healthy control under study. The positivity rate of HPV-11 among studied women with BC was 14.8% compared to 6.9% among the healthy control with an insignificant difference between the two groups ( $P=0.288$ ). The most affected age among BC women with

positive HPV-11 was less than 50 years with the highest positivity rate and significant association ( $P=0.007$ ).

Additionally, Human papillomavirus type 16 was identified with the highest positivity rate of 55.7% versus 3.4% the positivity rate of healthy control with a highly significant difference between the two groups ( $P=0.0001$ ). While the most affected age of BC women with positive HPV-16 was  $\geq 60$  years.

Moreover, the highest positivity rate of BC women with positive HPV-18 was 47.5% compared to 6.9% of the healthy control with a highly significant difference between the two groups ( $P=0.0001$ ). Additionally, the most affected age was less than 40 years with the highest positivity rate. High-risk Human papillomaviruses type 16 and type 18 were identified with the highest positivity rate among unmarried women. Human papillomaviruses under study were frequently found with the highest positivity rate among women with BC within the normal weight category with a significant association with HPV-16 only.

Additionally, the positivity rate of P53 overexpression was 65.6% among breast cancer women compared to the 6.9% positivity rate among the healthy control with highly significant differences ( $P=0.0001$ ). The serological levels of mutant P53 overexpression were found to be age-dependent ( $P= 0.029$ ). The Mean  $\pm$  SD was higher in the age range  $< 40$  years which was ( $633.1 \pm 208.0$ ) with significant association ( $P=0.029$ ). A higher, but insignificant association was found between P53 overexpression and oral contraceptives consumption ( $P=0.090$ ). Furthermore, the P53 was significantly associated with HPV-18 ( $P=0.001$ ), but not with HPV-11 or HPV-16.

In conclusion, the association of infectious rate of HPV types was generally high among Iraqi women with breast cancer. The highest infectious rate in order of frequency was HPV- 16, then HPV-18 and HPV-11. The most affected age of women with breast cancer was less than 50 years. P53 overexpression has a highly significant association with breast cancer, as well as with HPV-18 infection only

# Table of Contents

|  |            |
|--|------------|
| <b>SUMMARY .....</b>   | <b>I</b>   |
| <b>TABLE OF CONTENTS .....</b>                                       | <b>III</b> |
| <b>LIST OF TABLE .....</b>   | <b>VII</b> |
| <b>LIST OF FIGURES.....</b>  | <b>IX</b>  |
| <b>LIST OF ABBREVIATIONS .....</b>                                   | <b>X</b>   |
| <b>1. INTRODUCTION .....</b>   | <b>1</b>   |
| 1.2. AIM OF THE STUDY .....  | 3          |
| <b>2. LITERATURE REVIEW .....</b>                                    | <b>5</b>   |
| 2.1. DISCOVERY OF HUMAN PAPILLOMAVIRUS:.....                         | 5          |
| 2.2. HUMAN PAPILLOMAVIRUS CHARACTERISTICS: .....                     | 5          |
| 2.3. CLASSIFICATION OF THE VIRUS: .....                              | 6          |
| 2.4. HUMAN PAPILLOMAVIRUS TRANSMISSION:.....                         | 7          |
| 2.4.1. Non-sexual route: .....                                       | 8          |
| 2.4.2. Sexual transmission:.....                                     | 8          |
| 2.5. HUMAN PAPILLOMAVIRUS LIFE CYCLE.....                            | 9          |
| 2.6. CLINICAL MANIFESTATION OF HUMAN PAPILLOMAVIRUS.....             | 11         |
| 2.6.1. Skin lesions: .....   | 11         |
| 2.6.1.1. Benign skin lesions: .....                                  | 11         |
| 2.6.1.1.1 Common warts: .....  | 11         |
| 2.6.1.1.2. Filiform warts:.....                                      | 12         |
| 2.6.1.1.3. Plantar warts:.....                                       | 12         |
| 2.6.1.1.4. Flat warts:.....  | 12         |
| 2.6.1.1.5. Pigmented warts: .....                                    | 12         |
| 2.6.1.1.6. Epidermodysplasia verruciforme: .....                     | 12         |
| 2.6.1.2. Malignant skin lesions:.....                                | 13         |
| 2.6.1.2.1. Squamous cell carcinoma of the skin Squamous:.....        | 13         |
| 2.6.1.2.2. Bowen's disease: .....                                    | 13         |
| 2.6.1.2.3. Basal cell carcinoma of the skin:.....                    | 13         |
| 2.6.2. Mucosal lesions:.....   | 14         |
| 2.6.2.1. Benign mucosal lesions: .....                               | 14         |
| 2.6.2.1.1. Focal epithelial hyperplasia or Heck's disease:.....      | 14         |
| 2.6.2.1.2. Condyloma acuminata:.....                                 | 14         |
| 2.6.2.1.3. Bowenoid papulosis: .....                                 | 14         |
| 2.6.3. Malignant mucosal lesions:.....                               | 15         |
| 2.6.3.1. Queyrat's erythroplasia .....                               | 15         |
| 2.6.3.2. Vulvar cancer .....   | 15         |
| 2.6.3.3. Penile carcinoma: .....                                     | 15         |
| 2.6.3.4. Anal carcinoma .....  | 16         |
| 2.6.3.5. Cervical cancer.....  | 16         |
| 2.6.3.4. Oral and cervical cancer: .....                             | 16         |
| 2.6.3.5. Recurrent respiratory papillomatosis and lung cancer: ..... | 17         |
| 2.6.3.6. Breast cancer: .....  | 17         |
| 2.7. IMMUNE RESPONSE AGAINST HPV VIRUS: .....                        | 18         |
| 2.7.1. Innate immunity of HPV: .....                                 | 19         |
| 2.7.2. Adaptive immunity of HPV: .....                               | 21         |
| 2.7.3. Humoral immunity against HPV: .....                           | 21         |
| 2.7.4. Cellular immune response towards HPV: .....                   | 21         |
| 2.7.5. Human papillomavirus modulation of host immune system: .....  | 22         |
| 2.8. TP53 GENE: .....  | 23         |
| 2.8.1. TP53 gene associated with breast cancer: .....                | 23         |

|   |           |
|---|-----------|
| 2.9. ANATOMICAL STRUCTURE OF THE BREAST .....                     | 24        |
| 2.9.1. Breast cancer: .....                                       | 25        |
| 2.9.1.1. classification of breast cancer.....                     | 26        |
| 2.9.1.1.1. Histopathological types .....                          | 26        |
| 2.9.1.1.1.1. Ductal carcinoma in situ (DCIS): .....               | 27        |
| 2.9.1.1.1.2. Lobular Carcinoma in Situ (LCIS):.....               | 27        |
| 2.9.1.1.1.3. Invasive ductal carcinoma:.....                      | 27        |
| 2.9.1.1.1.4. Invasive lobular carcinoma: .....                    | 27        |
| 2.9.1.1.1.5. Medullary carcinoma:.....                            | 28        |
| 2.9. 1.1.1.6. Invasive micropapillary carcinoma .....             | 28        |
| 2.9.1.1.2. Grade .....  | 28        |
| 2.9.1.1.3. Stage.....   | 29        |
| 2.9.1.1.3.1. Tumour (T).....                                      | 29        |
| 2.9.1.1.3.2. Lymph node (N) .....                                 | 30        |
| 2.9.1.1.3.3. Metastasis (M).....                                  | 30        |
| 2.9.2. Breast cancer epidemiology .....                           | 30        |
| 2.9.3. Risk factor for the breast cancer.....                     | 31        |
| 2.9.4. Triple assessment in the diagnosis of breast cancer .....  | 32        |
| 2.10. LABORATORY DIAGNOSIS OF HUMAN PAPILLOMAVIRUS: .....         | 32        |
| 2.10.1. Lab diagnosis of non-genital warts: .....                 | 32        |
| 2.10.2. Lab diagnosis of genital warts:.....                      | 32        |
| 2.11. PROTECTION AGAINST HUMAN PAPILLOMA VIRUS INFECTION:.....    | 34        |
| 2.11.1. Vaccine component:.....                                   | 34        |
| 2.11.2. Immunogenicity and Vaccine Efficacy: .....                | 35        |
| 2.12. PREVALENCE OF HPV SUBTYPES WITH BREAST CANCER: .....        | 36        |
| <b>3. MATERIALS AND METHODS.....</b>                              | <b>40</b> |
| 3.1. MATERIALS: .....   | 40        |
| 3.1.1. Laboratory equipment: .....                                | 40        |
| 3.1.2. Laboratory appliances:.....                                | 40        |
| 3.1.3. Laboratory instruments: .....                              | 41        |
| 3.1.4. laboratory kits: .....                                     | 41        |
| 3.2. METHODS: .....   | 41        |
| 3.2.1. Study group: .....   | 41        |
| 3.2.1.1. Normal healthy women: .....                              | 42        |
| 3.2.1.2. Women with breast cancer: .....                          | 42        |
| 3.2.2. Sampling techniques: .....                                 | 43        |
| 3.2.3. Laboratory techniques:.....                                | 44        |
| 3.2.3.1. Determination of complete blood count: .....             | 44        |
| 3.2.3.1.1. Principle of the test:.....                            | 44        |
| 3.2.3.1.2. Automated procedure: .....                             | 44        |
| 3.2.3.2. Determination of Human papillomavirus -11 antigen: ..... | 44        |
| 3.2.3.2.1. Principle of the test:.....                            | 44        |
| 3.2.3.2.2. Laboratory Procedure:.....                             | 45        |
| 3.2.3.2.3. Interpretation of the result: .....                    | 46        |
| 3.2.3.3. Determination of Human papillomavirus -16 antigen: ..... | 46        |
| 3.2.3.3.1. Principle of the test:.....                            | 46        |
| 3.2.3.3.2. Laboratory Procedure:.....                             | 47        |
| 3.2.3.3.3. Interpretation of the result: .....                    | 48        |
| 3.2.3.4. Determination of Human papillomavirus -18 antigen: ..... | 48        |
| 3.2.3.4.1. Principle of the test:.....                            | 48        |
| 3.2.3.4.2. Laboratory Procedure:.....                             | 48        |
| 3.2.3.4.3. Interpretation of the result: .....                    | 49        |
| 3.2.3.5. Determination of Human P53 tumour protein:.....          | 50        |
| 3.2.3.5.1. Principle of the test:.....                            | 50        |
| 3.2.3.5.2. Laboratory Procedure: .....                            | 50        |
| 3.2.3.5.3. Interpretation of the result: .....                    | 52        |

|  |            |
|--|------------|
| 3.3. STATISTICAL ANALYSIS: .....   | 52         |
| <b>4. RESULTS.....</b>   | <b>53</b>  |
| 4.1. AGE AND BMI OF STUDY GROUPS:.....   | 53         |
| 4.2. ASSOCIATION BETWEEN DEMOGRAPHIC VARIABLES AND STUDY GROUPS:.....                            | 54         |
| 4.3. COMORBIDITIES:.....   | 55         |
| 4.4. MENSES: .....   | 55         |
| 4.5. THE EFFECT OF THE RISK FACTORS ON STUDY GROUPS: .....                                       | 56         |
| 4.5.1. Family history of malignancy: .....   | 56         |
| 4.5.2. Hormonal treatment: .....   | 56         |
| 4.6. CLINICOPATHOLOGICAL CHARACTERISTICS OF BREAST CANCER AMONG IRAQI WOMEN WITH BC:.....        | 57         |
| 4.7. DIAGNOSTIC TECHNIQUE VARIABLES AMONG IRAQI WOMEN WITH BREAST CANCER: .....                  | 58         |
| 4.8. POSITIVITY RATE OF HPVs AMONG STUDY GROUPS: .....   | 59         |
| 4.9. POSITIVITY RATE OF P53 OVEREXPRESSION AMONG STUDY GROUPS:.....                              | 60         |
| 4.10. HEMATOLOGICAL INDICES OF ASSOCIATED GROUPS: .....  | 60         |
| 4.10.1. ABO blood group:.....  | 60         |
| 4.10.2. complete blood count indices CBC: .....  | 61         |
| 4.11. SEROLOGICAL RESULTS: .....   | 62         |
| 4.11.1. Human papillomavirus type 11: .....  | 62         |
| 4.11.1.1. Association of demographic variables with HPV-11 among Iraqi women with BC:.....       | 62         |
| 4.11.1.2. Association of HPV-11 with comorbidities among Iraqi women with BC: .....              | 63         |
| 4.11.1.3. The effect of risk factors on Iraqi women with BC and positive HPV-11: .....           | 64         |
| 4.11.1.4. Clinicopathologic characteristic of BC among patients with positive HPV-11: .....      | 65         |
| 4.11.1.5. Diagnostic technique variables among Iraqi women with BC and positive HPV-11:.....     | 66         |
| 4.11.1.6. Association of HPV-11 with HPV-16, HPV-18 types and P53.....                           | 67         |
| 4.11.2. Human papillomavirus type 16: .....  | 68         |
| 4.11.2.1. Association of HPV-16 with demographic variables among Iraqi women with BC .....       | 68         |
| 4.11.2.2. Association of HPV-16 with comorbidities among Iraqi women with BC: .....              | 69         |
| 4.11.2.3. The effect of risk factors on Patients with BC and positive HPV-16: .....              | 70         |
| 4.11.2.4. Clinicopathological characteristic of BC among patients with positive HPV-16:.....     | 71         |
| 4.11.2.5. Diagnostic technique variables among breast cancer patients with positive HPV-16:..... | 72         |
| 4.11.2.6. Association of HPV-16 with HPV-11, HPV-18 types and P53.....                           | 73         |
| 4.11.3. Human papillomavirus type -18: .....   | 74         |
| 4.11.3.1 Association of HPV-18 with demographic variables among Iraqi women with BC: .....       | 74         |
| 4.11.3.2. Association of HPV-18 with comorbidities among Iraqi women with breast cancer:.....    | 75         |
| 4.11.3.3. The effect of risk factors on Iraqi women with breast cancer and positive HPV-18:..... | 76         |
| 4.11.3.4. Clinicopathologic characteristic of BC among patients with positive HPV-18: .....      | 77         |
| <b>5. DISCUSSION.....</b>  | <b>101</b> |
| 5.1. HUMAN PAPILLOMA VIRUSES POSAITIVITTY RATE: .....  | 102        |
| 5.1.1. Human papillomavirus type11 status: .....   | 105        |
| 5.1.2. Human papillomavirus type 16 status : .....   | 106        |
| 5.1.3. Human papillomavirus type 18 status : .....   | 108        |
| 5.2. P53 OVEREXPRESSION STATUS:.....   | 109        |
| 5.3. ASSOCIATION OF HPV WITH THE HISTOLOGICAL CLASSIFICATION OF THE BC :.....                    | 112        |
| 5.4. ASSOCIATED RISK FACTORS:.....   | 113        |
| 5.4.1. Body mass index: .....  | 113        |
| 5.4.2. Comorbidities: .....  | 114        |
| 5.4.3. Age distribution:.....  | 115        |
| 5.4.4. Marital status: .....   | 116        |
| 5.4.5. Family history of malignancy: .....   | 117        |
| 5.4.6. Using oral contraceptive:.....  | 117        |
| 5.4.7. Smoking: .....  | 118        |
| 5.5. PATHOLOGICAL ANALYSIS: .....  | 118        |
| 5.5.1. Diagnosis:.....   | 118        |
| 5.5.2. Diagnostic technique variables among Iraqi women with breast cancer:.....                 | 120        |
| 5.6. HAEMATOLOGICAL ANALYSIS: .....  | 120        |

|   |            |
|---|------------|
| <b>6. CONCLUSIONS AND RECOMMENDATIONS .....</b> | <b>122</b> |
| CONCLUSIONS .....                               | 122        |
| <b>REFERENCES .....</b>                         | <b>123</b> |
| <b>SUMMARY IN ARABIC.....</b>                   | <b>139</b> |
| <b>TITLE IN ARABIC.....</b>                     | <b>142</b> |

## List of Table

|   |    |
|---|----|
| Table 3-1: Laboratory equipment used in a current study. -----  | 36 |
| Table 3-2: Laboratory appliances used in the present study. -----                                     | 36 |
| Table 3-3: Laboratory instrument used in the present study. -----                                     | 37 |
| Table 3-4: Laboratory diagnostic Kits used in this study. -----                                       | 37 |
| Table 4-1: Distribution of age range and BMI of study group. -----                                    | 50 |
| Table 4-2: The effect of social factors on study groups. -----  | 50 |
| Table 4-3: Comorbidities among study groups. -----  | 51 |
| Table 4-4: Menses status among study groups. -----  | 52 |
| Table 4-5: Association of Family history of malignancy with breast cancer. ---                        | 52 |
| Table 4-6: Hormonal treatment and exposure to radiation. -----  | 53 |
| Table 4-7: Clinicopathological characteristics of breast cancer. -----                                | 54 |
| Table 4-8: Diagnostic technique variables among BC patients. -----                                    | 55 |
| Table 4-9: Association of HPVs understudy with study groups. -----                                    | 55 |
| Table 4-10: The association of P53 with study groups. -----   | 56 |
| Table 4-11: Distribution of ABO blood type among study groups. -----                                  | 57 |
| Table 4-12: Distribution of Complete blood counts with study groups. -----                            | 58 |
| Table 4-13: Distribution of age ranges among patients with HPV-11 and other variables. -----          | 59 |
| Table 4-14: The Association of HPV-11 with comorbidities among BC patients. -----                     | 60 |
| Table 4-15: The effect of risk factors on BC Patients with positive HPV-11. --                        | 61 |
| Table 4-16: Tumor characteristics among patients with HPV-11. -----                                   | 52 |
| Table 4-17: Diagnostic technique variables among patients with positive HPV-11. -----                 | 63 |
| Table 4-18: The association of HPV-11 with HPV-16, HPV-18 types and P53.                              | 64 |
| Table 4-19: Distribution of age ranges among patients with HPV-16 and other variables. -----          | 65 |
| Table 4-20: Association of HPV-16 with comorbidities among BC patients. ---                           | 66 |
| Table 4-21: The effect of risk factors on BC Patients with positive HPV-16. ---                       | 67 |
| Table 4-22: Tumor characteristic among patients with positive HPV-16. -----                           | 68 |
| Table 4-23: Diagnostic technique variables among patients with positive HPV-16. -----                 | 69 |
| Table 4-24: Association of HPV-16 with HPV-11, HPV-18 types and P53. ----                             | 70 |
| Table 4-25: Distribution of age ranges among patients with positive HPV-18 and other variables. ----- | 71 |
| Table 4-26: Association of HPV-18 with comorbidities among BC patients. ---                           | 72 |
| Table 4-27: The effect of risk factors on BC Patients with positive HPV-18. ---                       | 73 |

|   |    |
|---|----|
| Table 4-28: Tumor characteristic among patients with positive HPV-18. -----                                   | 74 |
| Table 4-29: Diagnostic technique Variables among patients with positive HPV-18. -----                         | 75 |
| Table 4-30: Association of HPV18 with HPV11, HPV16 types and P53. -----                                       | 76 |
| Table 4-31: The P53 positivity rate among breast cancer patients. -----                                       | 77 |
| Table 4-32: The association of P53 with comorbidities among Iraqi women with BC. -----                        | 78 |
| Table 4-33: The effect of risk factors on P53 overexpression among Iraqi women with BC. -----                 | 79 |
| Table 4-34: Association of tumour characteristics with P53 overexpression among Iraqi women with BC. -----    | 80 |
| Table 4-35: Association of P53 overexpression with diagnostic technique variables. -----                      | 81 |
| Table 4-36: Association of P53 with Human papillomavirus study types. -----                                   | 82 |
| Table 4-37: Association of demographic variables and histological grading among breast cancer patients. ----- | 83 |
| Table 4-38: Association of comorbidities with histological grading. -----                                     | 84 |
| Table 4-39: The effect of risk factors on histological grading. -----   | 85 |
| Table 4-40: Association of clinicopathological factors with histological grading. -----                       | 86 |
| Table 4-41: Association of diagnostic technique with histological grading. -----                              | 87 |
| Table 4-42: Association of HPVs study types and P53 with breast cancer grading. -----                         | 87 |
| Table 4-43: The distribution of complete blood count with different carcinoma diagnoses. -----                | 88 |
| Table 4-44: The distribution of complete blood count with different tumour grades. -----                      | 89 |
| Table 4-45: The distribution of complete blood count among HPV- 11. -----                                     | 90 |
| Table 4-46: The distribution of complete blood count among HPV- 16. -----                                     | 90 |
| Table 4-47: The distribution of complete blood count among HPV- 18. -----                                     | 91 |
| Table 4-48: Correlation of P53 titer with demographic variables among Iraqi women with BC. -----              | 92 |
| Table 4-49: Correlation of P53 titer with comorbidities among BC women. -----                                 | 93 |
| Table 4-50: Correlation of P53 titer with risk factor variables among BC patients. -----                      | 93 |
| Table 4-51: Correlation of P53 titer with clinicopathological variables among Iraqi women with BC. -----      | 95 |
| Table 4-52: Correlation of P53 titer with diagnostic procedures among Iraqi patients with BC. -----           | 96 |

## List of Figures

|   |    |
|---|----|
| Figure 2-1: Structure of Human papillomavirus. -----  | 5  |
| Figure 2-2: Human papillomavirus life cycle. -----  | 9  |
| Figure 2-3: Immune response against HPV. -----  | 17 |
| Figure 2-4: Role of Innate Immunity against Human Papillomavirus infections and<br>Effect of Adjuvants in Promoting Specific Immune Response. ----- | 18 |
| Figure 2-5: Anatomy of the breast. -----  | 23 |

## List of Abbreviations

| <b>Abbreviations</b>      | <b>Meaning</b>                          |
|---------------------------|---|
| BC                        | Breast Cancer BC                        |
| BCC                       | Basal Cell Carcinoma                    |
| BD                        | Bowen's Disease                         |
| CBC                       | Complete Blood Count                    |
| cSCC                      | cutaneous Squamous Cell Carcinoma       |
| DC                        | Dendritic Cells                         |
| DCIS                      | Ductal Carcinoma In Situ                |
| E1, E2, E4, E5, E6 and E7 | Early region proteins                   |
| EDTA                      | Ethylenediaminetetraacetic Acid         |
| EGFRs                     | Epidermal Growth Factor (EGF) Receptors |
| ELISA                     | Enzyme-Linked Immunosorbent Assay       |
| ER                        | Endoplasmic Reticulum                   |
| FDA                       | Food and Drug Administration            |
| FFPE                      | Formalin-Fixed Paraffin-Embedded tissue |
| FNA                       | Fine Needle Aspiration                  |
| HNCs                      | Head and Neck Cancers                   |
| HPV                       | Human Papillomavirus                    |
| HPV11                     | Human Papillomavirus type11             |
| HPV16                     | Human Papillomavirus type16             |
| HPV18                     | Human Papillomavirus type18             |
| HR                        | High - Risk                             |
| HRA                       | High-Resolution Anoscopy                |
| HRP                       | Horseradish Peroxidase                  |
| HSPGs                     | Heparin Sulphate Proteoglycans Receptor |

|           |  |
|-----------|--|
| IDC       | Invasive Ductal Carcinoma IDC              |
| IL-1      | Interleukin 1                              |
| ILC       | Invasive Lobular Carcinoma                 |
| IMPC      | Invasive Micropapillary Carcinoma          |
| L1 and L2 | Late region proteins                       |
| LC        | Langerhans Cells                           |
| LCIS      | Lobular Carcinoma In Situ                  |
| LNM       | Lymph Node Metastasis                      |
| LR        | Low-Risk                                   |
| LVI       | Lymph Vascular Invasion                    |
| M         | Metastasis                                 |
| MCH       | Mean Corpuscular Haemoglobin               |
| MCHC      | Mean Corpuscular Haemoglobin Concentration |
| MCV       | Mean Corpuscular Volume                    |
| MRI       | Magnetic Resonance Imaging                 |
| N         | Lymph Node                                 |
| NK        | Natural Killer Cells                       |
| NKT       | Natural Killer T Cells                     |
| NMSC      | Non-Melanoma Skin Cancer                   |
| OD        | Optical Density                            |
| P53       | Tumour suppressor protein                  |
| Pap       | Papanicolaou test                          |
| PCR       | Polymerase Chain Reaction                  |
| PML       | Promyelocytic Leukaemia                    |
| RDW       | Red Blood Cell Distribution Width          |
| RONs      | Reactive Oxygen-Nitrogen Species           |
| RRP       | Recurrent Respiratory Papillomatosis RRP   |

|       |   |
|-------|---|
| T     | Tumour  |
| TGF-β | Transforming Growth Factors - Beta                    |
| Th1   | T-helper cells type 1                                 |
| TNM   | Histopathological type, grade, stage (staging system) |
| VLPs  | Purified Virus-Like Particles                         |
| RB    | Retinoblastoma  |

# Chapter One

---

## 1. Introduction

Human papillomaviruses (HPVs) represent a large collection of viral types. *Papillomaviridae* is a family of non- envelope viruses whose members are called Human papillomavirus. More than 200 different types of human papillomavirus (HPV) have been identified to date. The papillomavirus was first identified at the beginning of the 20th century when it became clear that skin warts or papillomas could be transmitted between individuals by a filterable infectious agent (Christensen, 2016). In 1935, Francis Peyton Rous, who had previously demonstrated the existence of carcinogenic sarcomas in chickens, further showed that the papillomavirus can cause skin cancer in infected rabbits (Rous *et al.*, 2021). This was the first demonstration that the virus could cause cancer in mammals. Human papillomavirus is small, non-enveloped of 52-55 nm diameter with icosahedral shape. The viral capsid is not covered by a lipid membrane. The capsid protein is composed of 22 pentameric capsomers. The viral genome containing a single double-stranded DNA molecule with approximately 8000 base pairs (bp) which are bounded to cellular histones. The genome encodes eight major proteins, 6 located in the “early” region, proteins are E1, E2, E4, E5, E6 and E7 are regulatory in function. Two proteins are located in the “late” region L1 and L2 comprise the virus capsid required for virus transmission, spread and survival in the environment (Dcis *et al.*, 2019). Human papillomavirus can transmit by two routes, sexual route and non-sexual route, Two types of HPV (types 6 and 11) which induce benign genital warts or condylomata acuminate, they’re considered low-risk HPV because they don’t lead to cancer or other serious health problems. At least a dozen types of HPV can sometimes lead to cancer, though two (types 16 and 18) lead to the majority of cancer cases.

These are called high-risk HPV. Cervical cancer is most linked to HPV, but HPV can also cause cancer in the vulva, vagina, penis, anus, mouth, throat as well as breast

# Chapter One

---

cancer. It can also cause cancer in the back of the throat, including the base of the tongues and tonsils (called oropharyngeal cancer) (Bucchi, 2016).

Human papillomaviruses infection occurs at the basal cell layer of stratified squamous epithelial cells. Infection stimulates cellular proliferation in the epithelium and infected cells display a broad spectrum of changes, ranging from benign hyperplasia to dysplasia to invasive carcinoma. To effectively replicate, HPV must utilize the host cellular machinery. During the process, the viral protein product encoded by E6 binds to the p53 tumour suppressor gene product, which results in the premature degradation of the p53 protein (Mantovani and Banks, 2001). The E7 protein binds to a tumour suppressor protein—the retinoblastoma protein—and inhibits its function(Pang and Thierry, 2013). These protein products mediate much of the virus' oncogenic potential and their production represents a key difference between the low- and high-risk strains of HPV (Reed and Zazove, 2013).

Human papillomavirus infection may be latent, subclinical, or clinical. It may take the pathway of low viral-load infection without clinical disease, or high viral-load infection with clinical disease(Juckett and Hartman-Adams, 2010). HPV can cause a wide range of infections ranged from benign lesions to malignant infections.

Human Papillomaviruses (HPVs) are recognized as carcinogenic agents in breast cancer in humans (Salman *et al.*, 2017a). For many reasons, the relationship between HPV and breast cancer is imperative. The anatomy of the breast duct exposes the open duct to the external environment, which increases the risk of HPV infection. Most cases of breast cancer originate from the epithelium of the breast ducts, where hyperproliferation of ducts is followed by subsequent tumor progression to invasive ductal carcinoma *in situ*. Therefore, it is speculated that HPV virus particles can be transported from the original infection site in the genital area, enter through the nipple and infect the mammary duct. Later, this may be the cause of certain types of breast cancer (Sher *et al.*, 2020).

# Chapter One

---

## 1.2. Aim of the study

The present study was conducted to achieve the following goals:

1. Find an association between HPV and breast cancer among women who were histologically diagnosed as breast cancer among patients from Baghdad and different cities in Iraq by using an Enzyme-linked Immunosorbent Assay (ELISA kit)
2. Finding an association with tumour suppressor proteins P53 by using ELISA technique.